SI 1: Pathogen identification and ecology

The identity and ecology of the specific strains of pathogens found here are not well known, and thus all analyses and discussion are kept at genus level. However, here we use our best knowledge about the natural history and identify several of these specific strains.

Anaplasma – Except for one poor quality sequence, all *Anaplasma* sequences were 99-100% identical to *Anaplasma* sp. strain AnAj360 (GenBank accession AF497580.1). Unfortunately, this marker cannot accurately resolve species and the reference databases are quite confused. Among the closest named species was *Anaplasma platys* isolate 3ax1' (GenBank KJ659045.1), which differed by one to two nucleotides. *Anaplasma platys* causes canine anaplasmosis and is present in Kenya [1]. It is generally transmitted by the brown dog tick (*Rhipicephalus sanguineus*) which is regionally present. While this species of tick generally prefers large mammal hosts, it has also been known to parasitize small mammals in other systems [2,3]. *Anaplasma platys* has previously been detected in rodents and shrews as well as in ticks collected from them [4]. However, we emphasize that the ecology of *A. platys* may or may not be relevant to the strain identified here, given the lack of resolution in the database. Other species of *Anaplasma* present within the region in Kenya include *Anaplasma ovis* [5], and *Anaplasma marginale* [6].

Bartonella -. For *Bartonella*, we identified at least 17 major strains (some with substrains) in the 226 sequenced PCR products. *Bartonella* is an often rodent-associated pathogen whose vectors include ticks, sandflies, and fleas [7] and can infect a wide range

of mammals, including humans [8]. Many *Bartonella* species have been detected in Kenya, including *B. tribocorum, B. quenslandensis, B. rochalimae, B. grahamii, B. elizabethae,* and several other unidentified genotypes of *Bartonella* [9,10].

Borrelia – All *Borrelia* sequences had 100% nucleotide identity to *Borrelia burgdorferi*. However, flagellin is a conserved genetic region. There have been very few cases of Lyme disease (caused by *B. burgdorferi*) in Kenya, but two cases were reported in 2005 [11]. Other species of *Borrelia* reported regionally include *Borrelia duttonni*, *B. crocidurae*, and *B. recurrentis* (potentially just an ecotype of *B. duttonni*)[11-15], all of which use rodents as reservoir hosts. All of these latter species of *Borrelia* utilize soft ticks (primarily in the *Ornithodoros* genus) as vectors. Some of these species are known to cause relapsing fever in humans [14].

Theileria – We identified only one *Theileria* strain in this study. It was most closely related to *T. equi* (89% similarity). *Thieileria equi* causes equine piroplasmosis, which is known to affect zebras and donkeys in northern Kenya. It uses *Hyalomma* tick species as vectors [16]. Several species of *Hyalomma* ticks were found on small mammals in this study. However, 89% similarity is rather distant so it is unclear if this would be indicative of the ecology of this *Theileria* strain.

Hepatozoon - Two major strains, both with substrains, were found of *Hepatozoon*. This included one strain most closely related to *H. arygbor* and *H. erhardovae*, and the other most closely related to *H. felis* and *H. ursi* (Fig SI 2). *Hepatozoon* transmission, unlike

most vector borne pathogens, is completed by the ingestion of an ectoparasite (including ticks, fleas, mites, and lice) infected with *Hepatozoon* oocysts. Many *Hepatozoon* species have been found in Kenya; those genetically identified include *H. canis* and *H. felis* [18]

SI Table 1: Land use change effects on species richness and diversity across sites including the total number of species found across a land-use type, the average species richness per site, and the average Shannon diversity per site. Note, because sampling efforts were not equal across all land-use types, the total number of species captured should not be directly compared. Further analysis of the effects of land-use on species diversity can be found in [19]

Land Use	Total number of species	Average species richness	Average Shannon diversity
Conserved	25	5.22	1.10
Exclosure	18	7.83	1.38
Pastoral	17	4.24	1.17
Cropland	17	4.38	1.02

		Land use ef	Main effects				Interaction effects			
ANAPLASMA	AIC, df 300.6, 45			302.7, 45				304.2, 45		
		Ζ	SE	р	Ζ	SE	р	Ζ	SE	р
Intercept		-2.14	0.2	<0.0001	-1.79	1.12	0.12	-2.07	1.14	0.08
Land use	Cropland	1.78	0.29	<0.0001	1.8	0.29	0	1.41	2.11	0.51
	Exclosure	0.51	0.3	0.1	0.5	0.3	0.1	-1.78	1.62	0.28
	Pastoral	0.65	0.32	<0.01	0.64	0.23	< 0.01	5.98	2.07	0.01
Rainfall					0	0.002	0.75	0.000	0.002	0.99
Interactions	Rainfall: Cropland							0.000	0.003	0.89
	Rainfall:Exclosure							0.004	0.003	0.18
	Rainfall:Pastoral							-0.090	0.004	0.01
BORRELIA	AIC, df	268.9, 45			270.4, 45			268.7, 45		
		Z	SE	р	Ζ	SE	р	Ζ	SE	р
Intercept		-2.24	0.15	<0.0001	-2.49	0.77	< 0.01	-2.3	0.87	0.01
Land use	Cropland	1.13	0.28	<0.001	1.11	0.28	< 0.001	0.07	1.91	0.97
	Exclosure	0.65	0.28	0.02	0.67	0.28	0.02	1.98	1.39	0.16
	Pastoral	-0.48	0.23	0.04	-0.49	0.23	0.04	-1.89	1.93	0.33
Rainfall					0	0	0.74	0.000	0.000	0.93
Interactions	Rainfall: Cro	pland						0.002	0.000	0.59
	Rainfall:Exc	losure						0.000	0.000	0.35
	Rainfall:Past	oral						0.000	0.000	0.47

SI Table 2: Effects of land use, rainfall and their interaction on the five focal pathogens

		Land use e		All main e	ffects		Interaction effects			
BARTONELLA	AIC, df 220.8, 45			227.2, 45			236.6, 45			
		Ζ	SE	р	Ζ	SE	р	Ζ	SE	р
Intercept		0.37	0.11	<0.01	-0.34	0.59	0.57	-0.93	0.72	0.2
Land use	Cropland	0.52	0.22	0.02	0.48	0.23	0.04	1.97	1.64	0.24
	Exclosure	0.66	0.22	<0.01	0.66	0.23	0.01	-1.98	1.25	0.14
	Pastoral	-0.15	0.18	0.41	-0.1	0.19	0.6	3.63	1.66	0.04
Rainfall					0.001	0.001	0.24	0.002	0.001	0.09
Interactions	Rainfall: Cro	opland						0.000	0.002	0.37
	Rainfall:Exclosure						0.005		0.06	
	Rainfall:Pas	storal						0.007	0.003	0.03
THEILERIA	AIC, df				395.2, 45			284.6, 45		
		Ζ	SE	р	Ζ	SE	р	Ζ	SE	р
Intercept					395.2	2.29	<0.0001	-13.12	0.99	< 0.001
Land use	Cropland				3.1	0.38	<0.0001	7.52	1.98	< 0.01
	Exclosure				86	0.4	0.04	7.18	1.53	< 0.001
	Pastoral				0.37	0.3	0.23	4.01	1.95	0.05
Rainfall					0.01	0	<0.01	0.016	0.002	< 0.0001
Interactions	Rainfall: Cr	opland						-0.008	0.003	<0.01
	Rainfall:Ex	closure						-0.012	0.003	<0.001
	Rainfall:Past	oral						-0.006	0.004	0.884

HEPATOZOON	Land use effects			Main effects only				Interaction effects			
	AIC, df	283.6, 45			285.7, 45						
		Z	SE	р	Ζ	SE	р	Ζ	SE	р	
Intercept		-0.87	0.16	<0.0001	-0.89	0.84	0.29	-0.82	0.96	0.4	
Land use	Cropland	0.61	0.3	0.05	0.61	0.31	0.06	0.34	2.14	0.88	
	Exclosure	1.13	1.13	<0.001	1.13	0.30	< 0.001	-1.18	1.59	0.46	
	Pastoral	-0.35	-0.35	0.17	-0.35	0.25	0.18	0.6	2.17	0.78	
Rainfall					0.000	0.001	0.99	-0.001	0.002	0.95	
Interactions	Rainfall: Cr	opland						0.000	0.003	0.90	
	Rainfall:Exclosure							0.004	0.003	0.17	
	Rainfall:Pas	toral						-0.002	0.004	0.66	

ANAPLASMA			Z	SE	р
	Intercept		-2.35	0.27	<0.0001
	Driver	Diversity	0.39	0.21	0.07
		Density	2.86	0.64	<0.001
BORRELIA			Ζ	SE	р
	Intercept		-3.1	0.25	<0.0001
	Driver	Diversity	0.31	0.19	0.11
		Density	3.36	0.6	<0.0001
BARTONELLA			Ζ	SE	р
	Intercept		-0.69	0.17	<0.001
	Driver	Diversity	0.27	0.13	0.04
		Density	3.85	0.41	<0.0001
THEILERIA			Ζ	SE	р
	Intercept		-5.9	0.59	<0.0001
	Driver	Diversity	0.34	0.35	0.34
		Density	6.29	1.05	<0.0001
HEPATAZOON			Ζ	SE	р
	Intercept		-2.38	0.21	<0.0001
	Driver	Diversity	0.64	0.16	<0.0001
		Density	3.84	0.5	<0.0001

SI Table 3: Effects of diversity and density on abundance of infected animals for each pathogen

FLEAS		Ma	ain effects only	Interaction effects fects only model					
	AIC, df	289.18, 45			302				
			Ζ	SE	р	Z	SE	p	
	Intercept		1.944	0.88	0.03	-1.33	1.11	0.23	
	Landuse	Cropland	-0.31	0.3	0.31	4.88	2.16	0.04	
		Exclosure	0.47	0.3	0.12	-2.014	1.64	0.22	
		Pastoral	0.15	0.25	0.55	8.04	2.16	<0.01	
	Rainfall		0.002	0.001	0.18	0.007	0.002	<0.01	
	Interactions	Rainfall:Croplan	d			-0.010	0.003	0.02	
		Rainfall:Exclosu	re in the second s			0.005	0.003	0.12	
		Rainfall:Pastora	I			-0.014	0.004	<0.001	

SI Table 4: Effects of land use, rainfall and their interaction on the flea abundance

Fig SI 1: GARLI tree depicting the 17 major strains of *Bartonella* identified across the rodent samples.

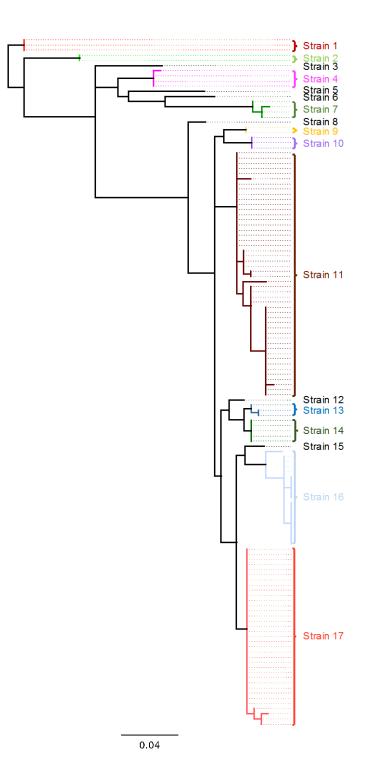
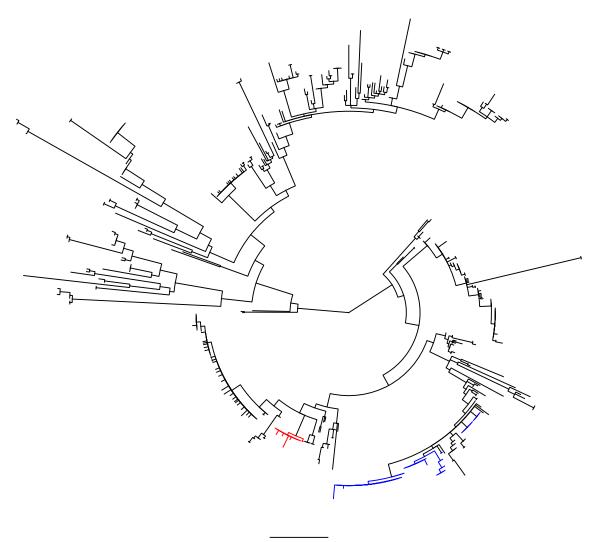


Fig SI 2: GARLI tree depicting the phylogenetic position of the two major *Hepatozoon* lineages found in this study in comparison to other strains found on GenBank (in black). Red: strain most similar to *H.felis* and *H. ursi*; Blue: strain most similar to *H. ayorgbor* and *H. erhardovae.*



0.09

Fig SI 3: For three pathogens – *Anaplasma* (A), *Theileria* (B), and *Hepatozoon* (C) there was modest support for a positive relationship between diversity of small mammals at a site and the number of infected animals per site for any pathogen, after correction for spatial autocorrelation. The other two pathogens showed no relationship. Using site level species richness instead of Shannon diversity produces qualitatively similar (but stronger) effects.

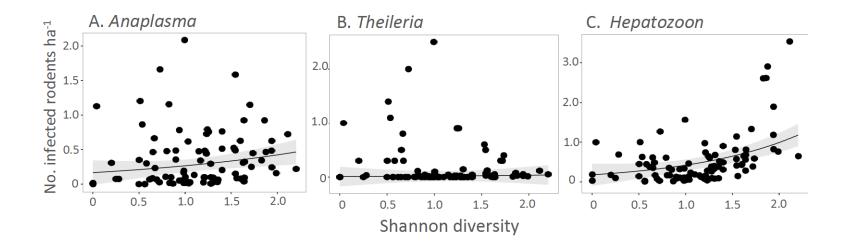


Fig SI 4: There was no significant relationship between site level Shannon diversity and site level density of small mammals in any land use type.

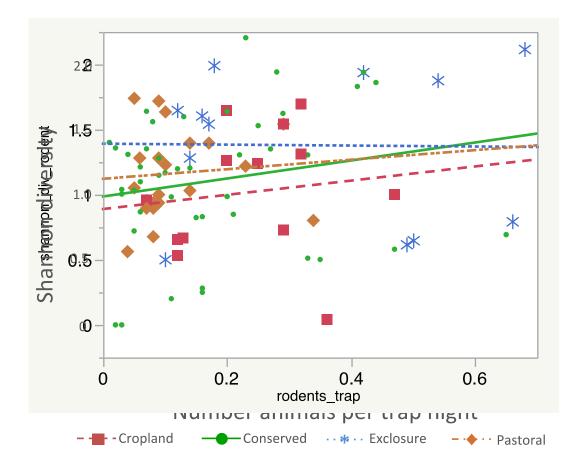


Fig SI 5: Density of infected small mammals was positively correlated with the abundance (measured as number of animals per trap night) of small mammals; however this effect was stronger in cropland sites where there were a higher proportion of infected animals.

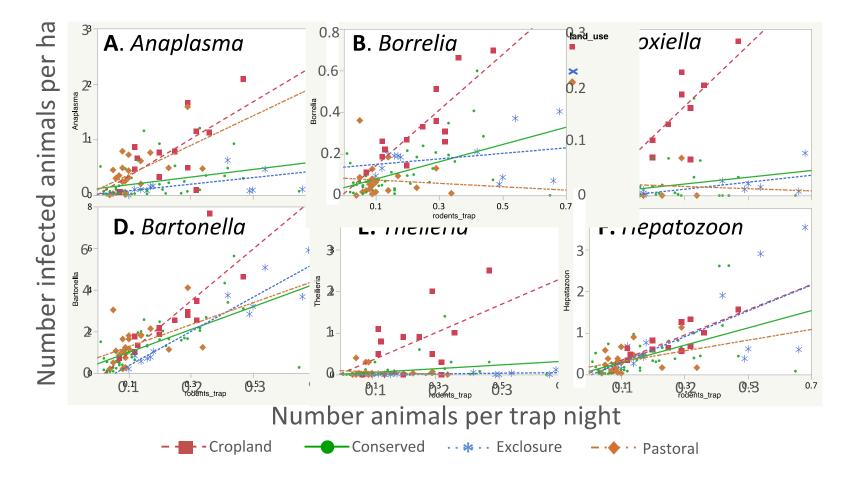
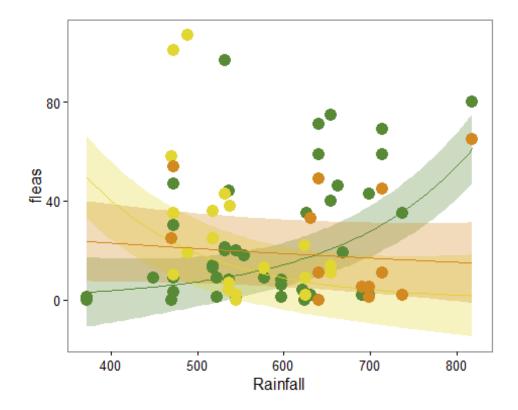


Fig SI 6: Flea abundance results closely paralleled those seen for *Bartonella*, with the number of fleas decreasing with rainfall in pastoral sites but increasing with rainfall in conserved sites.



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