#### PROKARYOTES



# Draft Genome Sequence of Uncultured Upland Soil Cluster *Gammaproteobacteria* Gives Molecular Insights into High-Affinity Methanotrophy

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**ABSTRACT** Aerated soils form the second largest sink for atmospheric CH<sub>4</sub>. A nearcomplete genome of uncultured upland soil cluster *Gammaproteobacteria* that oxidize CH<sub>4</sub> at <2.5 ppmv was obtained from incubated Antarctic mineral cryosols. This first genome of high-affinity methanotrophs can help resolve the mysteries about their phylogenetic affiliation and metabolic potential.

A erated soils remove 4 to 6% of  $CH_4$  from the atmosphere (1) as a result of high-affinity methanotrophy that has been demonstrated by the detection of the high-affinity form of particulate methane monooxygenase, encoded by *pmo* genes (2–6). In the absence of isolates and genomes, phylogenetic identity and physiology of atmospheric methane-oxidizing bacteria have remained a mystery (7). Here, we report a near-complete genome of uncultured upland soil cluster *Gammaproteobacteria* (USC $\gamma$ ) obtained from incubated mineral cryosols (pH 8.51) collected from Taylor Dry Valley, Antarctica. These samples exhibited CH<sub>4</sub> oxidation when incubated with 2.5 ppmv at 4°C and 10°C, and were found to contain USC $\gamma$ -like *pmo*A genes (8).

DNA was extracted using PowerSoil total DNA/RNA isolation kit (MO BIO Laboratories, Inc., CA), purified and concentrated using Microcon centrifugal filters 100 (EMD Millipore Corp., MA). Four metagenomic libraries were prepared using the PrepX DNA library kit and the automated Apollo  $324^{TM}$  robotic system (WaferGen Bio-systems, Inc., CA), pooled at equal molar amount and sequenced on Illumina HiSeq 2500 (Illumina Inc., CA). Paired-end reads (2 × 100 nucleotides [nt]; mean insert size of 350 bp) were quality filtered (9). High-quality reads (average 75.7 ± 7.5 million reads per sample) were coassembled using IDBA\_UD v1.1.1 (default settings with precorrection) (10). Protein-encoding genes were predicted using Prodigal v2.6.1 (11) and searched for *pmo*A genes. All scaffolds were subjected to binning by an interactive approach (12) and MetaBAT (13).

A high-quality genome bin containing a *pmo*A gene 100% identical to one of our USC $\gamma$ -like *pmo*A clone sequences was identified. Reads mapped to the scaffolds in this genome bin were reassembled using IDBA\_UD (default settings). CheckM (14) determined that it was 89.99 to 91.9% complete with 0 to 0.09% contamination and 0% strain heterogeneity. Annotation was performed using Prokka v1.12-beta (15), BLAST (16), and KEGG Automatic Annotation Server v2.1 (17). A single copy of twenty 30S and thirty-one 50S ribosomal proteins, one 5S rRNA operon, a single copy of 16S and 23S rRNA gene fragments, 38 tRNA genes, and 3,012 coding sequences were retrieved. The

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488 nt-long 16S rRNA gene shared 99% identity to uncultured bacteria (JQ684308, HM445440, and DQ823229) and 94% identity to *Thioalkalivibrio* (NR\_074692) and *Ectothiorhodospira* (NR\_125567) of *Chromatiales*, which is phylogenetically closely related to *Methylococcales* but includes no methanotrophs.

The USC $\gamma$  draft genome contained the complete operon of *pmo*CAB. The *pmo*A gene shared 72% identity with *Methylocaldum szegediense* (*Methylocacales*). Genes encoding methanol dehydrogenase and accessory proteins, enzymes for the tetrahydrofolate and tetrahydromethanopterin-linked C1 transfer pathways, and two formate dehydrogenases were identified. Although gene encoding formaldehyde dehydrogenase (FALDH) was missing, genes for pyrroloquinoline quinone (PQQ) synthesis were identified, indicative of the potential use of PQQ-dependent FALDH. As with the genomes of other gammaproteobacterial methanotrophs, the USC $\gamma$  draft genome encodes all essential genes for a complete serine biosynthesis pathway for formaldehyde assimilation. Genes encoding enzymes for nitrogen metabolism were also identified. Manual annotation and phylogenetic and comparative analyses are in progress to pinpoint the phylogenetic affiliation of USC $\gamma$  and to elucidate differences in the metabolic potential between high- and low-affinity methanotrophs.

**Accession number(s).** This whole-genome shotgun project and the USC $\gamma$  draft genome have been deposited at DDBJ/ENA/GenBank under the accession no. MUGK00000000. The version described in this paper is version MUGK01000000.

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#### REFERENCES

- Kirschke S, Bousquet P, Ciais P, Saunois M, Canadell JG, Dlugokencky EJ, Bergamaschi P, Bergmann D, Blake DR, Bruhwiler L, Cameron-Smith P, Castaldi SC, Chevallier F, Feng L, Fraser A, Heimann M, Hodson EL, Houweling S, Josse B, Fraser PJ, Krummel PB, Lamarque J-F, Langenfelds RL, Le Quéré C, Naik V, O'Doherty S, Palmer PI, Pison I, Plummer D, Poulter B, Prinn RG, Rigby M, Ringeval B, Santini M, Schmidt M, Shindell DT, Simpson IJ, Spahni R, Steele LP, Strode SA, Sudo K, Szopa S, van der Werf GR, Voulgarakis A, van Weele M, Weiss RF, Williams JE, Zeng G. 2013. Three decades of global methane sources and sinks. Nat Geosci 6:813–823.
- Holmes AJ, Roslev P, McDonald IR, Iversen N, Henriksen K, Murrell JC. 1999. Characterization of methanotrophic bacterial populations in soils showing atmospheric methane uptake. Appl Environ Microbiol 65: 3312–3318.
- Knief C, Lipski A, Dunfield PF. 2003. Diversity and activity of methanotrophic bacteria in different upland soils. Appl Environ Microbiol 69:6703–6714. https://doi.org/10.1128/AEM.69.11.6703-6714.2003.
- Kolb S. 2009. The quest for atmospheric methane oxidizers in forest soils. Environ Microbiol Rep 1:336–346. https://doi.org/10.1111/j.1758-2229 .2009.00047.x.
- Nauer PA, Dam B, Liesack W, Zeyer J, Schroth MH. 2012. Activity and diversity of methane-oxidizing bacteria in glacier forefields on siliceous and calcareous bedrock. Biogeosci Discuss 9:1259–1298. https://doi.org/ 10.5194/bgd-9-1259-2012.
- 6. Lau MCY, Stackhouse BT, Layton AC, Chauhan A, Vishnivetskaya TA,

Chourey K, Mykytczuk NCS, Bennett PC, Lamarche-Gagnon G, Burton N, Ronholm J, Pollard WH, Omelon CR, Medvigy DM, Hettich RL, Pfiffner SM, Whyte LG, Onstott TC. 2015. An active atmospheric methane sink in high Arctic mineral cryosols. ISME J 9:1880–1891. https://doi.org/10.1038/ ismej.2015.13.

- Knief C. 2015. Diversity and habitat preferences of cultivated and uncultivated aerobic methanotrophic bacteria evaluated based on *pmoA* as molecular marker. Front Microbiol 6:1346. https://doi.org/10.3389/fmicb .2015.01346.
- Lau MCY, Edwards CR, Onstott TC. 2016. SPaMOB can eat atmospheric methane in Antarctica. Abstr 2016. AGU fall meeting, abstr B53C-0538, San Franscisco, CA.
- Lau MCY, Kieft TL, Kuloyo O, Linage-alvarez B, van Heerden E, Lindsay MR, Magnabosco C, Wang W, Wiggins JB, Guo L, Perlman DH, Kyin S, Shwe HH, Harris RL, Oh Y, Yi MJ, Purtschert R, Slater GF, Ono S, Wei S, Li L, Sherwood Lollar B, Onstott TC. 2016. An oligotrophic deep-subsurface community dependent on syntrophy is dominated by sulfur-driven autotrophic denitrifiers. Proc Natl Acad Sci U S A 113:E7927–E7936. https://doi.org/10.1073/pnas.1612244113.
- Peng Y, Leung HCM, Yiu SM, Chin FYL. 2012. IDBA-UD: a *de novo* assembler for single-cell and metagenomic sequencing data with highly uneven depth. Bioinformatics 28:1420–1428. https://doi.org/10.1093/ bioinformatics/bts174.
- Hyatt D, Chen GL, Locascio PF, Land ML, Larimer FW, Hauser LJ. 2010. Prodigal: prokaryotic gene recognition and translation initiation site

identification. BMC Bioinformatics 11:119. https://doi.org/10.1186/1471 -2105-11-119.

- Albertsen M, Hugenholtz P, Skarshewski A, Nielsen KL, Tyson GW, Nielsen PH. 2013. Genome sequences of rare, uncultured bacteria obtained by differential coverage binning of multiple metagenomes. Nat Biotechnol 31:533–538. https://doi.org/10.1038/nbt.2579.
- Kang DD, Froula J, Egan R, Wang Z. 2015. MetaBAT, an efficient tool for accurately reconstructing single genomes from complex microbial communities. PeerJ 3:e1165. https://doi.org/10.7717/peerj.1165.
- 14. Parks DH, Imelfort M, Skennerton CT, Hugenholtz P, Tyson GW. 2015. CheckM: assessing the quality of microbial genomes recovered from

isolates, single cells, and metagenomes. Genome Res 25:1043–1055. https://doi.org/10.1101/gr.186072.114.

- Seemann T. 2014. Prokka: rapid prokaryotic genome annotation. Bioinformatics 30:2068–2069. https://doi.org/10.1093/bioinfor matics/btu153.
- Camacho C, Coulouris G, Avagyan V, Ma N, Papadopoulos J, Bealer K, Madden TL. 2009. Blast+: architecture and applications. BMC Bioinformatics 10:421. https://doi.org/10.1186/1471-2105-10-421.
- Moriya Y, Itoh M, Okuda S, Yoshizawa AC, Kanehisa M. 2007. KAAS: an automatic genome annotation and pathway reconstruction server. Nucleic Acids Res 35:W182–W185. https://doi.org/10.1093/nar/gkm321.



## Erratum for Edwards et al., "Draft Genome Sequence of Uncultured Upland Soil Cluster *Gammaproteobacteria* Gives Molecular Insights into High-Affinity Methanotrophy"

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Volume 5, no. 17, e00047-17, 2017, https://doi.org/10.1128/genomeA.00047-17. Page 2: Lines 11–14 should read as follows: "...Unlike the genomes of other gammaproteobacterial methanotrophs, the USC $\gamma$  draft genome contains nearly all essential genes for a complete serine biosynthesis pathway for formaldehyde assimilation. However, the enzymes unique to the typical RUMP pathway, hexulose-6-phosphate synthase and hexulose phosphate isomerase, were not detected...."

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